Heat shock proteins in three related *Drosophila* species belonging to the *obscura* group

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Abstract. The effect of heat shock on protein synthesis in three related Drosophila species belonging to the obscura group was analyzed on SDS-acrylamide gels. Four major heat shock proteins (hsps) were found in these species, in which synthesis reaches a maximum at 34 °C. Although the higher molecular weight proteins are conserved, differences in size were found for the small hsps in these species. By means of in situ hybridization using D. melanogaster probes for the small hsp genes, it was inferred that the small hsp genes of the obscura group species are clustered at the 27A locus in all three species.

Key words. Drosophila-related species; obscura group; heat shock; hsps.

All organisms respond to temperature shock by inducing the synthesis of a specific set of proteins called heat shock proteins, hsps (reviewed elsewhere¹⁻⁵). Three major types of hsps are found in most organisms: a hsp with a molecular weight of 70 kDa (hsp70), a larger hsp of 80-90 kDa, and a group of small hsps with MW between 15 kDa and 30 kDa. Comparing the hsps in Drosophila a high degree of evolutionary conservation is found in the high MW proteins. Hsps of 82,000 M_r and 70,000 M_r have been reported in all species studied⁶⁻⁹. Small hsps are recognized as a heterogeneous group where the size and number of the proteins vary between species. In order to check whether the small hsp family shows a different pattern even in related species, we compared the heat shock patterns of protein synthesis in three sibling Drosophila species of the obscura group: Drosophila subobscura, D. guanche and D. madeirensis.

The strains used in this study were H 271 of *D. subobscura*, TF 2 of *D. guanche*, and Md 1 of *D. madeirensis*. All three are isofemale strains deriving from individuals captured in Finland, the Canary Archipelago and Madeira Island, respectively. Individuals of the three species synchronized at the beginning of prepupa formation and kept at 19 ± 1 °C, were exposed to temperatures of 31 °C, 34 °C or 37 °C over a period of 30 min. Controls were obtained from synchronized prepupae kept at 19 °C during the treatment period. Experimental design was that described in Pascual and de Frutos¹⁰. Samples were electrophoresed in 9-12% SDS-acrylamide gels according to Laemmli's method¹¹ and fluorography of the gels was carried out following the Bonner and Laskey's procedure¹².

The effect of heat shock on protein synthesis was similar in the three species analyzed. Synthesis reached a maximum at 34 °C (fig. 1a). A dramatic decrease in protein synthesis was found at 37 °C which is the optimum

temperature to switch on the heat shock machinery in *Drosophila melanogaster*. As can be seen in this figure, all three species produced four particularly abundant hsps that were not synthesized in significant amounts under control conditions. The induced high MW proteins show identical electrophoretic mobility in all three species, and correspond to hsp82 and hsp70 found in *D. melanogaster*. Hsp70 is also the main product synthesized after heat shock in the *obscura* group species. The table summarizes the results obtained in these species and compares them with that of *D. melanogaster*.

Four of the small hsps have been described in D. melanogaster^{6,7}, and can be separated into the doublets 28-26 kDa and 23-22 kDa by electrophoresis in low concentration acrylamide gels (fig. 1b). D. subobscura, D. guanche and D. madeirensis always showed two small hsps under the same experimental conditions. The two small proteins found in D. subobscura were previously reported by Pascual and de Frutos¹⁰. As can be seen in the table, the sizes of the small proteins were not the same in the three related species. Comparison of the amino acid sequences of the D. melanogaster small hsps indicates a large region with a high degree of sequence homology and two heterologous regions which account for the variable size of these proteins (reviewed in Craig¹³). The size differences in the *obscura* small hsps may be for similar reasons.

In situ hybridization using *D. melanogaster* probes for the four small hsp genes (88.5, 88.6)¹⁴, gave positive signals with both probes at the 27A locus on the J polytene chromosome of the three *obscura* group species. The 88.5 and 88.6 probes contain, respectively, the hsp28-hsp23 and hsp26-hsps22 genes from the 67B locus of *D. melanogaster*. As an example, figure 2 shows the in situ results using the 88.5 probe. Chromosomes of the three species were prepared following Atherton

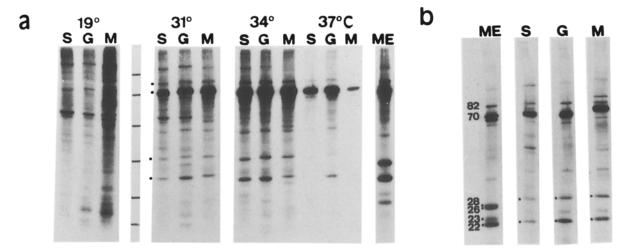


Figure 1. Heat shock proteins in three obscura group Drosophila species. a Proteins were analyzed before heat shock at control temperature, and after heat treatments in D. subobscura (S), D. guanche (G) and D. madeirensis (M). ME, D. melanogaster has pattern at 37 °C. Molecular weights are indicated by short black lines which correspond to 92,500, 66,200, 45,000, 31,000, 21,500 and 14,500 daltons. b Electrophoretic distribution on 9% polyacrylamide gels of has in these species. The D. melanogaster has pattern is indicated in kDa.

and Gall's procedure¹⁵. Probes were radiolabeled with ³H-dCTP (1',2',5-³H dCTP, Amersham, sp. act. 68 Ci/mmol) following the oligolabeling method described by Feinberg and Vogelstein¹⁶. Prehybridization, hybridization and subsequent washes were performed at high temperature as described by Pardue¹⁷.

From the in situ results, it was inferred that, as in D. melanogaster, genes encoding the small hsps are orga-

nized in a closed linkage group in all three obscura group species and probably came from one ancestral gene. During the evolution of Drosophila species, fewer changes must have taken place in the genes encoding hsps of high MW than in the genes of the small hsps, which show a wide range of size and number of proteins among the different species found today.

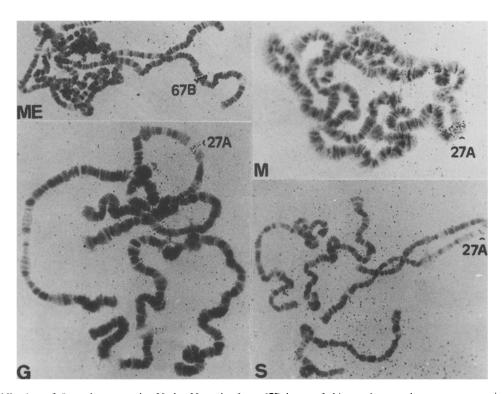


Figure 2. Hybridization of *D. melanogaster* hsp28-hsp23 probe from 67B locus of this species, to *obscura* group species polytene chromosomes. It shows the uniqueness of the hybridization site at the 27A locus on the J chromosome of these species. ME, *D. melanogaster*; G, D. guanche; M, D. madeirensis; S, D. subobscura.

Comparison between hsp patterns of the obscura group species with that of D. melanogaster. The small hsp apparent MW of the obscura group species are indicated in daltons

hsp family	D. melanogaster	D. subobscura	D. guanche	D. madeirensis
hsp82	+	+	+	+
hsp70 small hsps	+	+	+	+
hsp32	-	+ (32,000)	+ (33,000)	+ (33,000)
rsp28	+		<u> </u>	
sp26	+	_	_	<u> </u>
nsp23	+	+ (23,500)	+ (23,400)	+ (24,200)
nsp22	+		-	-

^{+,} presence; -, absence.

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- 1 Schlesinger, M. J., Tissières, A., and Ashburner, M., eds. Heat Shock Proteins: from Bacteria to Man. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY 1982.
- 2 Neidhardt, F. C., VanBogelen, R. A., and Vaughn, V., A. Rev. Genet. 18 (1984) 295.
- 3 Nover, L., ed. Heat Shock Response of Eukaryotic Cells. Springer-Verlag, Heidelberg 1984.
- 4 Lindquist, S., and Craig, E. A., A. Rev. Genet. 22 (1988) 631. 5 Nagao, R. T., Kimpel, J. A., and Key, J. L., in: Genomic Responses to Environmental Stress, p. 235. Ed. J. S. Scandalios.
- Academic Press, Inc., New York 1990. 6 Tissières, A., Mitchell, H. K., and Tracy, U. M., J. molec. Biol. 84 (1974) 389.

- 7 Lewis, M., Helmsing, P. J., and Ashburner, M., Proc. natl Acad. Sci. USA 72 (1975) 3604.
- 8 Sinibaldi, R. M., and Storti, R. V., Biochem. Genet. 20 (1982)
- 9 Blackman, R. K., and Meselson, M., J. molec. Biol. 188 (1986)
- 10 Pascual, L., and de Frutos, R., Chromosoma 97 (1988) 164.
- 11 Laemmli, U. K., Nature 277 (1970) 680.
- 12 Bonner, W. M., and Laskey, R. A., Eur. J. Biochem. 46 (1974)
- 13 Craig, E. A., CRC Crit. Rev. Biochem. 18 (1985) 239.
- 14 Corces, V., Holmgren, R., Freund, R., Morimoto, R., and Meselson, M., Proc. natl Acad. Sci. USA 77 (1980) 5390.
- 15 Atherton, D., and Gall, J., Dros. Inform. Serv. 49 (1972) 311.
- 16 Feinberg, A. P., and Vogelstein, B., Analyt. Biochem. 132 (1983) 6.
- 17 Pardue, M. L., in: Drosophila, a Practical Approach, p. 111. Ed. D. B. Roberts. IRL Press, Oxford, Washington, DC 1986.